



INFLUENCE OF DIFFERENT COLOURS OF LIGHTS ON DEVELOPMENTAL STAGES OF RED FLOUR BEETLE (*TRIBOLIUM CASTANEUM*) AND ADULTS PHOTOTAXIS BEHAVIOR

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ABSTRACT

Red flour beetle (*Tribolium castaneum*) is the major insect pest of store grain, and its single larvae can attack 88 grains during its life which leads to a considerable loss of quality and viability of grain. Therefore, to manage this insect pest present research was designed to check the effects of different colours of lights on the developmental stages of red flour beetle and their attraction on lights. Six (White, Yellow, Red, Green, Blue and Black) colours of light were used in the present experiment at $30\pm 2^{\circ}\text{C}$ and $60\%\pm 10\%$ relative humidity (RH). Results revealed that a longer incubation period (4.17 ± 0.18 , less hatching ($78.66\pm 0.50\%$) and highest mortality ($21.34\pm 1.11\%$) of *T. castaneum* eggs were observed on blue light, while a short incubation period (3.03 ± 0.51), highest hatching ($89.99\pm 1.92\%$) and lowest mortality ($10.01\pm 0.22\%$) were observed on white colour of light. However, a longer duration (32.01 ± 0.15 , 12.33 ± 0.31) and maximum (28.10 ± 2.50 , 14.66 ± 0.52) mortality of *T. castaneum* was found in larval and pupae stages on blue colour of light, while a short duration (21.27 ± 0.27 , 7.33 ± 0.13) and minimum (8.30 ± 2.5 , 8.00 ± 0.65) larval and pupal mortality (%) were seen by red and yellow colour of lights. Similarly, an adult's longevity significantly reduced (48.33 ± 1.76) when beetles were exposed on blue light, whereas a significant (62.33 ± 1.66) increase in adult longevity was found on white colour of light. A highest (33.33 ± 0.56) attraction of red flour beetle was found on red colour of light, whereas, a lowest attraction was observed on blue (6.00 ± 0.85) and black (10.00 ± 1.55) colours of lights.

Keywords: developmental period, hatching percentage, light attraction, mortality, *Tribolium castaneum*

INTRODUCTION

Stored grains are broken by numerous store grain insect pests such as mites, weevil, beetles and moths, resulting in quantitative and qualitative losses (Rajendran and sriranjini, 2008). Store grain insect pests contribute to contamination in grains commodities through the occurrence of insect body fragments, excretions and dead insects. Red flour beetle (*Tribolium castaneum*) is the major insect pest of store grain that causes both quantitative and qualitative losses which spoil the flour by feeding, and it is a very common pest in flour mills, warehouses and grocery stores (Garcia *et al.*, 2005). It is most economically key pest and has worldwide distribution (Padin *et al.*, 2002). A single larva of *T. castaneum* can attack 88 grains during its life which leads to a considerable loss of quality and viability of grain (Atanasov, 1978). Karunakaran *et al.* (2004) examined that infestation reason by red flour

beetle in unprotected kernel stored for nine months reduced germination virtually completely and augmented visually broken kernels from 9 to 39%. Therefore, controlling the *T. castaneum* relies on the use of fumigation and conventional insecticides, such as chlorfluazuron and methyl bromide (Kim and Lee, 2013). Use of fumigation and chemical to secure stored product commodities by insect infestation and contamination while their continuously uses leads to problems of unpleasant residues and developed resistance in certain insect species (Huang and Subramanyam 2004; Khattak and Malik 1979); Salam and Shakoori 1990). Until now, fumigation is the most effective method to manage these pests, but still has serious limitations, Such as fumigants do not penetrate some commodities in sufficient concentration to control some pests. However, in few instances, chemical treatments may have lethal influence on the grains or may leave adverse residues. Therefore, there is need to search for another pest management tactic for *T. castaneum*.

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Non-chemical control practices are attractive since they neither leave chemical residues in the grains and commodity nor do they cause resistance in insect pests (Padin *et al.*, 2002). Amongst the non-chemical coordination, lights effects have been used to manage many insects pests of stored grains such as the Indian meal moth and cigarette beetle population (Sambaraju and Phillips, 2008). However, dark light and light of incandescent bulbs have short electric effectiveness and wide wavelengths; therefore, more efficient light source such as light emitting diodes (LEDs) have been used to control insects through light traps (Cohnstaedt *et al.*, 2008; Lee *et al.* 2008). LEDs light traps have been broadly used to the management of *Liriomyza trifolii* and *Trialeurodes vaporariorum* insect pest (Kim and Lee, 2013). LEDs are most efficient because of their small size, light weight, low electric consumption, adjustable light intensity, long life time and capability to select specific wavelength (Tamulaitis *et al.*, 2005; Yeh and Chung, 2009). Hassan and Khan (1998) reported that light can be used against stored product pests by the direct treatment of the commodities that provide a residue-free process of pest control, because lights do not cause residues in grains and also no significantly change the quality of the stored food material or stored kernels. Keeping in view, the store grain product of Pakistan is under threat due to red flour beetle infestations; present study was carried out to determine the effect of different colour of lights on life cycle parameters and adults phototactic behavior of red flour beetle *in vitro*.

MATERIALS AND METHODS

Culture of Red flour beetle

The experiment was conducted in the Department of Entomology, Lasbela University of Agriculture, Water and Marine Sciences, (LUAWMS), Uthal, Balochistan. Red flour beetles (*Tribolium castaneum*) were collected from Uthal godowns and they were cultured in plastic boxes on wheat grain flour under laboratory conditions at $30\pm 2^{\circ}\text{C}$ and $60\%\pm 10\%$ RH. The light source was purchased from science, international laboratory in Karachi. Six (White, Yellow, Red, Green, Blue and Black) different colour of lights were used in present study. Lights were used in the tope center of cardboard box with the size of (30W x 30L x 30H). During experiment it covered with black cloth. Each light was separately used in each cardboard box.

Plastic boxes were used for insects rearing and covered with cotton yarn. Wheat flour was provided as a diet in whole experiment. All experiments were replicated three times. Three hundred fresh eggs were collected and exposed on different light colours in petri dishes (7 cm diameter) to check the hatching (%) and their incubation periods. After hatching, 200 young larvae were collected and released on different light colours in plastic boxes (3 x 3 x 4 cm) covered with cotton yarn to observe the mortality (%) and larval developmental period. One hundred, one day old pupae were collected and exposed to different light colours in petri dishes to find out the adult emergence (%), and pupal days. After emergence 50 adults were collected and exposed to different lights to observe the adult longevity until the death of adults. The phototactic behavior of adults *T. castaneum* was also investigated. Transparent plastic pipe was used to find the phototactic behavior of adults *T. castaneum*. Five feet transparent plastic pipe was crossed in the middle of 6 light cardboard boxes. Transparent plastic pipe was banned at its one side with cotton net and closed with rubber and other hole was left open for insect entrance. Hundred adults of *T. castaneum* were collected randomly and exposed to different colour of lights to observe the adults phototactic behavior of *T. castaneum*. The light attraction data were recorded after 24 hours.

Statistical analysis

Data was analyzed with the help of SPSS (SPSS Inc., Chicago, IL, US) software and One Way Analysis of Variance (ANOVA) was used for data analysis and means were compared with Tukey test at $P<0.05$.

RESULTS

A significant ($P<0.05$) hatching (%) of *T. castaneum* eggs was observed on white colour of light followed by yellow, red, green and black colour of light. While a minimum hatching percentage was observed on blue colour of light. No significant difference was observed among white, yellow, red, green and black colour of light except blue light (Figure 1).

A significant increased ($P<0.05$) in mortality (%) of *T. castaneum* eggs was found on blue colour of light, however a significant decreased percentage was noticed on white colour of light, although no significant different were seen among white, yellow, red, green and black colour light (Figure 2).

A maximum incubation period of *T. castaneum* eggs was recorded in blue colour followed by red, green, black and yellow colour light. However, minimum incubation period of eggs was observed in white color. Only significant ($P<0.05$) difference was noticed on blue colour light as contrasted with other colours of lights. No differences were seen by white, yellow, red, green and black light as mentioned in Figure 3.

A longest larval developmental duration of *T. castaneum* was recorded on blue colour followed by black, green, white and yellow colour. While a minimum larval development duration was observed on red colour. Statistically, a significant ($P<0.05$) difference was noticed on blue, black and green light as compared with white, yellow and red colours (Figure 4).

Larval mortality (%) of *T. castaneum* significantly ($P<0.05$) differed when larvae were exposed on blue and black colour of lights as compared to other colours of lights. While no significant changes were recorded in larval mortality on white, yellow, red and green colour of lights (Figure 5).

A longer duration of *T. castaneum* pupal period was recorded on blue colour of light, whereas a short duration of pupal period was observed on yellow colour of light. Only significant ($P<0.05$) difference was found on blue colour of light as compared with others light colours in pupal period of *T. castaneum*, however, no difference was observed in pupal period in white, yellow, red, green and black colour of lights (Figure 6).

A mortality (%) at pupal stage of *T. castaneum* was increased ($P<0.05$) in pupae exposed to blue colour of light as compared to other colours, however a significant decreased of *T. castaneum* mortality (%) at pupal stage was noticed on red colour of lights. But no difference was observed among white, yellow, red, green and black colour of lights (Figure 7).

A significantly ($P<0.05$) increased adults longevity of *T. castaneum* was observed on white, yellow, red and green colour of lights, however a significant decrease adult longevity was observed on blue and black colour of lights. But no significant variation was observed among white, yellow, green and red colours of lights on adult's longevity of *T. castaneum* (Figure 8).

A red colour of light was found more attractive to A highest significant ($P<0.05$) lights attraction on adults stage of *T. castaneum* were found on red colour, while a lowest light

attraction was observed on blue and black colour. Although no difference was seen among white, yellow and green colour for the attraction of *T. castaneum* adults (Figure 9).

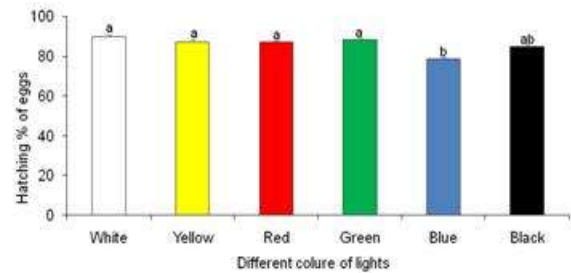


Figure 1. Influence of different colours of lights on the hatching (%) of *T. castaneum* eggs. A and B indicating the differences ($P<0.05$) among the groups

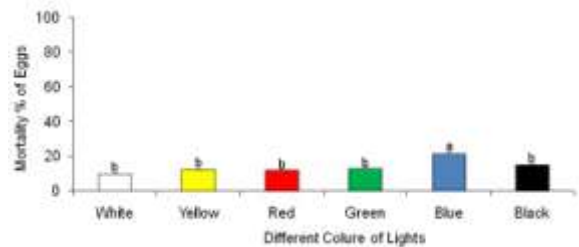


Figure 2. Influence of different colours of lights on the mortality (%) of *T. castaneum* eggs. A and B indicating the differences ($P<0.05$) among the groups

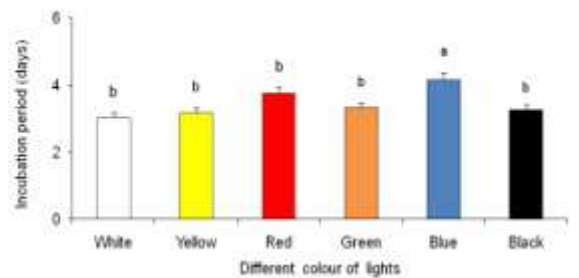


Figure 3. Influence of different colours of lights on the incubation period of *T. castaneum* eggs. A and B indicating the differences ($P<0.05$) among the groups

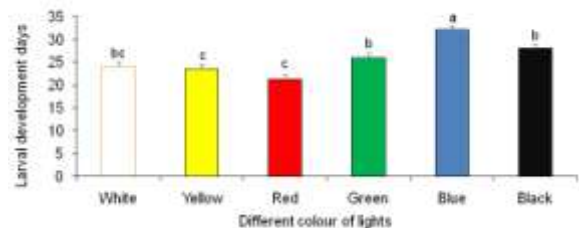


Figure 4. Influence of different colours of lights on the larval developmental days of *T. castaneum*. A, B and C indicating the differences ($P<0.05$) among the groups

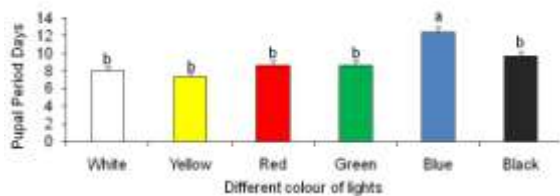


Figure 5. Influence of different colours of lights on the larval mortality (%) of *T. castaneum*. A, B and C indicating the differences ($P<0.05$) among the groups

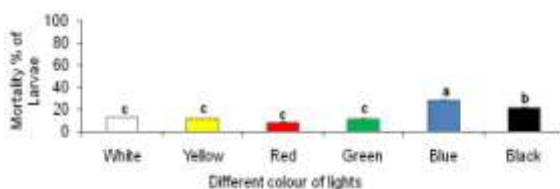


Figure 6. Influence of different colours of lights on the pupal developmental days of *T. castaneum*. A and B indicating the differences ($P<0.05$) among the groups

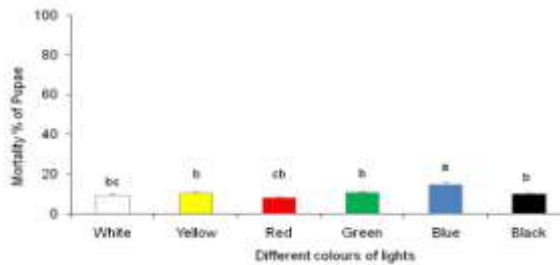


Figure 7. Different colour lights influence on the mortality (%) of *T. castaneum* pupae. . A, B and C indicating the differences ($P<0.05$) among the groups

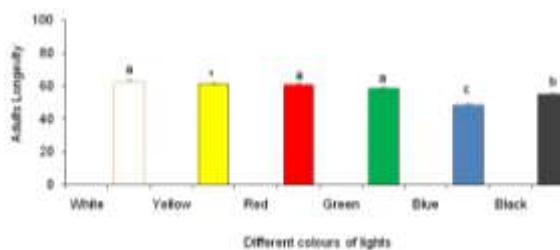


Figure 8. Influence of different colours of lights on the adults longevity of *T. castaneum*. . A, B and C indicating the differences ($P<0.05$) among the groups

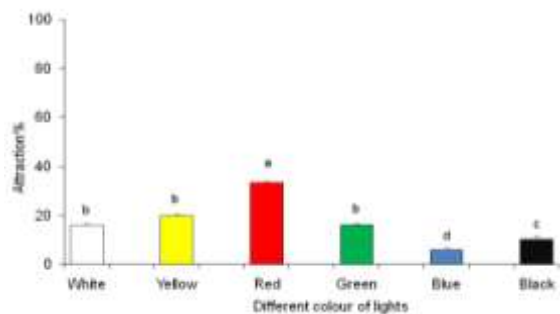


Figure 9. Adults Attraction of *T. castaneum* on different colour of lights. . A, B, C and D indicating the differences ($P<0.05$) among the groups

DISCUSSION

Various colour of lights have significantly effect on orientation, locomotion, feeding, mating, ovipositor, adult emergence, and the development of insects (Khan *et al.*, 1998). In present study we examined the influence of different colour of light on life history parameters and photo tactic behavior of red flour beetle *in vitro* and we observed that the less hatching ($78.66\pm 0.50\%$) and longer (4.17 ± 0.18), incubation periods of eggs were found on blue light colour as compared with white, yellow, green, red and black colour lights. These Findings indicated that growth of embryo might be inhibited by the energy of blue light. A similarly results also were reported by Hori *et al.* (2014); Hori and Suzuki, (2017); Shibuya *et al.* (2018). They reported that blue light has negative impact on the eggs of *D. melanogaste* and *Galerucella griseascens*. However a highest hatching (%) of eggs and lowest incubation period was recorded on white light. Agreed with the finding of Wang *et al.* (2013) who examined the highest eggs fecundity and fertility of bug *Orius sauteri* on white light.

Wang *et al.* (2013); Tariq *et al.* (2017) reported that blue light decrease the speed of metamorphosis of *Orius sauteri* and *B. dorsalis* and interruption in the growth of larvae to reach them at adult stage. Zhang *et al.* (2011); Ali *et al.* (2016) reported that black light enhance the larval period of *H. armigera* and *M. separate*. Similar results also were found in present study, and we observed longer (32.01 ± 0.15 , 28.03 ± 0.57), period of red flour beetle larvae on blue and black lights. While a short larval days (21.27 ± 0.27), and minimum mortality ($8.30\pm 2.5\%$) of red flour beetle was observed on red light, finding of red flour beetle on red lights regarding short periods of larvae and mortality supporting with the results of Sheribha *et al.* (2010), who also found the less mortality and decline in the average larvae periods of *T. castaneum* on red light. The accurate mechanism of reaction to colour lights was not known, however it was assumed that different lights of colour produced harmful effects on the metabolism of grubs. Such metabolic changes have been observed by Narayan *et al.* (1959); Vaidya *et al.* (1974); Khan *et al.* (1998). Using of different colour light traps methods incorporating colored lights is also promising mechanism in suppressing the pest population of red flour beetle in any infested stored grains godowns.

A maximum pupal day (12.33 ± 0.31) was noticed on blue light, whereas a minimum pupal

day (7.33 ± 0.13), was found on yellow light. Present results are supporting with the findings of Wang *et al.* (2013) who found that *Orius sauteri* took long time to reach at adult stage on blue light and less time were seen on green, yellow and white light for adults emergence. A highest mortality ($14.66 \pm 0.52\%$), at pupal stage was observed on blue light, however a lowest mortality ($8.00 \pm 0.65\%$), at pupal stage were observed on red light. similar consequence also were observed by Hori *et al.* (2014) because they has been observed the highest mortality of *D. melanogaster*, *C. pipiens molestus* and *T. confusum* at pupal stages on blue lights. However, the lowest mortality (%) at pupal stage of *T. castaneum* on red light are supporting with the findings of Sheribha *et al.* (2010) who found the maximum adults emergence percentage of *T. confusum* on red lights.

In present results regarding adult's longevity of *T. castaneum* a significant decrease adult's longevity of red flour beetle was noticed when it's exposed on blue light and black light as compared with other lights. Present findings are agreed with the findings of Sheeba *et al.* (2000), they reported that lights reduce the adult's life span in *D. melanogaster* and also agree with the investigation of Zhang *et al.* (2011); Ali *et al.* (2016) exposed that black light decrease the adult's longevity and reduce the fecundity of *H. armigera* and *M. separata*. Further our results are similar with the results of Hori *et al.* (2014); Hori and Suzuki (2017), who reported that blue light, has negative effect on different developmental stages of *Drosophila melanogaster* and declined their adult's life span. Specific species and developmental stage specific photoreceptive part might be connected with the dangerous impact of blue and black light. Reactive oxygen species (ROS) might contribute to spoil caused by blue and black light. Therefore, the insect body is injured by ROS, which causes the reduction of adult's longevity. These results are indicating that blue light and black light could suppress the next generation of *T. castaneum*.

Red colour light was found more attractive to the adults of red flour beetles as compared with others lights. Previously, it was reported that blue light (84.3%) was the most attractive to *Sitophilus oryzae* (L.), the rice weevil, followed by green, and UV LEDs lights (Jeon *et al.*, 2012). However, green and the blue LEDs 1.3 and 1.5 times more attractive than the others lights. This consequence is change from our findings, where blue colour was undesirable for

T. castaneum. These changes may be due to the insect's species and *T. castaneum* depend on the luminance specific wavelength and light exposure time. Semeao *et al.* (2011) found that factors such as light intensity and contrast with colour background may be impact the level of adults captured. Peitsch *et al.* (1992) Briscoe and Chittka (2001) investigated that Insects typically have photoreceptor that react to ultraviolet, blue and green lights. Furthermore, Bees of several species have natural preference for UV-blue and blue flowers (Giurfa *et al.*, 1995); Raine *et al.*, 2006). A number of insects have blue, green and UV photoreceptor in their eyes (Brisco and chittka, 2001). Vaishampayan *et al.* (1975) reported a phototactic response of *Trialeurodes vaporariorum* (westwood) to yellow, green and UV illuminance. Photoreceptor of *T. castaneum* adults has a single long wavelength opsin-across the entire retina (Jackowska *et al.*, 2007). Adults showed highest attraction on red light may be the reason of long wavelength-opsin of retina. On the basis of our findings, it was suggested that, 24 hours exposure of red light to *T. castaneum* was the most suitable in attracting *T. castaneum* adults for protecting stored foods/grains.

CONCLUSION

The present results revealed that different colours of lights have ability to disturb the physiological activities of *T. castaneum*. Colours may be considerable aspects of integrated pest managements alongside the other insects control application. Presents findings are essential towards providing baseline information for established pest control technique using various colours of lights. Useful pest control could be realized by clarifying the most lethal colour of lights of the targeted developmental stages such as eggs and pupae could be targeted due to unmovable stages. Larvae and adult insects that live in limited space could also be targeted, such as sanitary insect pest or stored grain insect's pest. Use of different colour lights for pest control, we must first identify the effective colour of each developmental stages for each targeted species. Furthermore, this study will provide important information on insect photobiology for researchers and extension workers.

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AUTHOR'S CONTRIBUTION

N. Ahmed: Research conducted
A. Ali: Experimental designed
S. A. Memon: Paraphrasing
T. K. Qambrani: Paraphrasing
G. Khaliq: Manuscript reviewed

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